

# Fungi found on *Helianthus annuus* in Finland

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Twenty species of fungi were identified in moist chamber cultures of diseased organs of *Helianthus annuus* L. Severe infection by *Botrytis cinerea* Nacca & Balbis and *Sclerotinia sclerotiorum* (Lib.) de Bary was demonstrated, and these species together with *Fusarium avenaceum* (Fr.) Sacc., *F. equiseti* (Corda) Sacc., *F. oxysporum* Schlecht. and *F. sambucinum* Fuckel form a disease complex which is very destructive on sunflower stands in Finland. These fungi have not been reported earlier on *Helianthus annuus* in Finland.

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## Materials and methods

The material for this study was provided by the author Tulisalo from trials with a sunflower (*Helianthus annuus* L.) variety (Garlic 64, Issanka, Peredovik) at the Agricultural Research Centre, in Vantaa, Tikkurila, Finland (Grid 27°E:6688:392). The samples were taken at random from apparently diseased capitula (18 samples), stems (15) and leaves (5), which were in moderately poor condition. Brownish or yellowish lesions occurred on the lamina or petioles of the leaves, and on the stems, which could break off at ground level. Most of the capitula had brownish lesions and soft rot, showing conidia of *Botrytis cinerea* or white mycelium and black sclerotia of *Sclerotinia sclerotiorum*. The samples were taken on August 6 and September 24, 1979.

The mycological studies were carried out in the Institute of Plant Pathology of the Agricultural Research Centre, Tikkurila. In the analyses two methods were used: 1) In the filter paper method about 1-cm<sup>3</sup> pieces of diseased plant were placed in Petri dishes (14 cm in diam.), in which there were one cotton filter and two filter papers, wetted with distilled water (20 ml). The plant pieces in the Petri dishes were about 1 cm apart, never touching each other. During the first two weeks the dishes were kept in room temperature (about +24°C) in diffuse daylight and for the following two weeks in +5°C in the dark. 2) In the agar method pieces of plants were disinfected with absolute alcohol for two minutes, dried and flamed. They were then placed on potato dextrose agar (PDA, Difco) in Petri dishes (9 cm in diam.) in the same way as in the filter paper method and kept in

diffuse daylight all the time. The fungi on the agar were determined after one and three weeks, and the cultures were kept continuously in room temperature. Three parallel Petri dish cultures were grown from each sample: one on filter paper and two on agar.

The identification of the species was facilitated by establishing separate agar cultures of the fungi, especially those of the genera *Botrytis*, *Fusarium* and *Sclerotinia*. Selected, dried specimens from agar cultures were deposited in the University herbarium (H).

The fungi were determined mainly according to von Arx (1981), Booth (1971; the *Fusarium* species), Domsch et al. (1980) and Ellis (1971).

## Results and discussion

In the present study 20 species were determined; five could be identified only to the genera. Most of the fungi that developed are common saprophytes on many kinds of plant material and in soil (see Domsch et al. 1980, Ellis 1971). Investigations showed that there were three severe pathogens: *Botrytis cinerea*, *Fusarium* spp. and *Sclerotinia sclerotiorum* (Table 1). *B. cinerea* and *S. sclerotiorum* caused especially extensive and severe infections on the sunflowers.

*S. sclerotiorum*, which is a major pathogen of the sunflower in sunflower-growing areas (Zimmer & Hoes 1978), chiefly destroyed the capitula, causing head rot, and the stems, causing basal rot, but was never found in the leaves. *Sclerotinia* first caused soft rot, then conspicuous external white mycelium formed and later the sclerotia developed. The fungus occurred only in the mycelial stage. On PDA the mycelium was white, and black sclerotia were formed.

*B. cinerea* was common on capitula and stems. *B. cinerea* and *S. sclerotiorum* almost always occurred together in capitula, while *Fusarium* was seldom found in the same sample. In seven cases, *B. cinerea* was found alone, without *Sclerotinia* or *Fusarium*, which shows that this fungus can infect sunflowers independently as well as developing as a secondary parasite. *B. cinerea* was very common in the stem bases, where it often occurred together with *Fusarium*, but only once with *S. sclerotiorum*. On PDA *B. cinerea* formed black sclerotia, which were smaller than those of *S. sclerotiorum*. The culture was grayish and there were numerous conidiophores with conidia. *B. cinerea* is not considered a serious disease of the sunflower elsewhere, and is reported to occur when wet weather delays the harvest (Zimmer & Hoes 1978).

*Fusarium* species were very common on rotted heads and stems. *Fusarium* species have not generally been recognized as serious sunflower pathogens (cf. Zimmer & Hoes 1978). *F. oxysporum* has been isolated from the sunflower in Argentina (Sackston 1957), and Orellana (1971) has reported that *Fusarium* causes wilting in the sunflower in Texas. In the present study, however, *Fusarium* species frequently occurred in capitula, stems and leaves, especially in rotten stem bases. The most common *Fusarium* species were *F. equiseti*, *F. oxysporum*, *F. sambucinum* and *F. avenaceum*. The species *F. culmorum* and *F. solani* were isolated much more rarely (Table 1).

Together with *Botrytis cinerea* and *Sclerotinia sclerotiorum* the species of *Fusarium* form a disease complex which is very destructive on sunflower crops in Finland, at least when ripening and harvesting have been delayed. The other major pathogens of the sunflower, *Puccinia helianthi* Schw. and *Verticillium dahliae* Klebahn, were not found (Zimmer & Hoes 1978).

The present fungus species have not been reported earlier growing on *Helianthus annuus* in Finland. The

sunflower is rare as a weed and is not yet grown as an oil crop in this country, but is not uncommon as an ornamental.

Table 1. Fungi developed on *Helianthus annuus* in moist chambers.

	capitula stems leaves		
Number of samples	18	15	5
Number of cultures	54	45	15
Fungi in cultures	%	%	%
Zygomycetes			
<i>Mucor</i> sp.	13	31	27
<i>Rhizopus</i> sp.	0	33	53
Ascomycetes			
<i>Sclerotinia sclerotiorum</i> (Lib.) de Bary	50	13	0
Deuteromycetes			
<i>Acremonium murorum</i> (Corda)			
W. Gams	0	0	13
<i>Acremonium</i> sp.	22	15	0
<i>Alternaria alternata</i> (Fr.) Keissler	28	0	60
<i>Arthrotrichum superba</i> Corda	0	11	0
<i>Aspergillus</i> sp.	0	7	53
<i>Botrytis cinerea</i> Nocca & Balbis	72	80	0
<i>Cladosporium cladosporioides</i> (Fres.) de Vries	6	0	20
<i>C. herbarum</i> (Pers.) Gray			
<i>Cylindrocarpum destructans</i> (Zinssm.) Scholten	0	4	0
<i>Fusarium avenaceum</i> (Fr.) Sacc.	0	27	20
<i>F. culmorum</i> (W.G.Sm.) Sacc.	0	4	0
<i>F. equiseti</i> (Corda) Sacc.	6	35	40
<i>F. oxysporum</i> Schlecht.	0	27	47
<i>F. sambucinum</i> Fuckel	0	40	27
<i>F. solani</i> (Mart.) Sacc.	0	9	0
<i>Fusarium</i> sp.	11	7	27
<i>Gliocladium roseum</i> Bainier	9	13	33
<i>Harzia acremoniioides</i> (Harz) Cost.	0	0	20
<i>Trichoderma viride</i> Gray	0	9	0
<i>Trichothecium roseum</i> (Pers.) Gray	0	7	0
<i>Ulocladium consortiale</i> (Thüm.) E. Simmons	9	33	27
<i>Verticillium tenerum</i> (Pers.) Link	4	20	0
<i>Verticillium</i> sp.	11	7	0
Unidentified fungi	6	11	13

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